# Journal of

# Gastroenterology and Hepatology Research

Online Submissions: http://www.ghrnet.org/index./joghr/doi:10.6051/j.issn.2224-3992.2014.03.399

Journal of GHR 2014 June 21 3(6): 1103-1107 ISSN 2224-3992 (print) ISSN 2224-6509 (online)

ORIGINAL ARTICLE

# **Does Colonic Transit Time Affect Colonic pH?**

### Ali Abbas, Gregory E Wilding, Michael D Sitrin

Ali Abbas, Michael D Sitrin, Department of Medicine, Division of Gastroenterology, Hepatology, and Nutrition, University at Buffalo, Buffalo, New York, the United States

Gregory E Wilding, Department of Biostatistics, University at Buffalo, Buffalo, New York, the United States

Correspondence to: Michael D Sitrin, MD, Department of Medicine, Division of Gastroenterology, Hepatology, and Nutrition, 462 Grider, Rm. 132-7, University at Buffalo, Buffalo, New York, the United States

Email: mdsitrin@buffalo.edu

Telephone: +1-716-898-5487 Fax: +1-716-898-5876 Received: December 21, 2013 Revised: April 9, 2014

Accepted: April 16, 2014 Published online: June 21, 2014

### **ABSTRACT**

**AIM**: To assess the effect of colonic transit time (CTT) on pH in the proximal and distal colon.

**METHODS**: CTT and pH in different regions of the colon were assessed using a wireless capsule manometry system. Healthy controls and patients with chronic constipation based on Rome II criteria were studied. Mean pH was determined in time quartiles of colonic transit, in the first 15 minutes after entry of the capsule into the cecum, and in the last 15 minutes prior to capsule exit from the body and correlated with CTT.

**RESULTS**: There was a weak, but significant correlation between CTT and pH in the distal colon (R=0.20, p=0.01 in the 15 minutes prior to body exit, and R=0.22, p=0.01 in the fourth quartile of CTT). No correlations between CTT and pH were found in the proximal colon. Patients with slow transit constipation tended to have a greater rise in pH from the proximal to the distal colon than those with normal transit constipation or healthy controls.

**CONCLUSION**: Longer CTT is associated with an increase in pH in the distal, but not proximal colon. The higher pH in the distal colon in slow transit constipation may affect important physiological and pathological processes.

© 2014 ACT. All rights reserved.

**Key words:** Gastrointestinal motility; Hydrogen ion concentration; Colon

Abbas A, Wilding GE, Sitrin MD. Does Colonic Transit Time Affect Colonic pH? *Journal of Gastroenterology and Hepatology Research* 2014; 3(6): 1103-1107 Available from: URL: http://www.ghrnet.org/index.php/joghr/article/view/735

### INTRODUCTION

Variation in intraluminal colonic pH has potentially important implications for colonic drug delivery<sup>[1-7]</sup>, alteration of colonic flora<sup>[8-12]</sup>, colorectal cancer carcinogenesis<sup>[13-15]</sup>, and regulation of colonic transport processes<sup>[16,17]</sup>. The colonic luminal pH progressively rises from the cecum to the rectum<sup>[18-21]</sup>. The major determinants of colonic pH are gut bacterial metabolism and colonocyte transport processes. There is a drop in pH by approximately one unit at the ileocecal transition due to generation of short chain fatty acids (SCFA) by colonic bacteria acting on dietary carbohydrates and fiber<sup>[22-26]</sup>. The uptake of SCFA by colonocytes along with luminal bicarbonate secretion contributes to the rise in pH in the distal colon<sup>[23,27]</sup>. Colonic luminal pH has been studied using a stationary pH electrode introduced by colonoscopy<sup>[18]</sup> or by a free falling radiotelemetry capsule<sup>[19,20]</sup>.

Few studies have looked at the effect of colonic transit time (CTT) on colonic pH<sup>[6,28,29]</sup>. In a small study by Lewis et al of 13 healthy subjects, senna, bran, and loperamide were used to alter gut transit. A positive correlation between whole gut transit estimated using fecal markers and distal colonic pH was noted<sup>[28]</sup>. The bran, however, may have also altered the colon pH by increasing the generation of short chain fatty acids. Thomas et al studied 40 patients, 20 healthy controls and 20 with gallstones, and found a positive correlation between CTT, estimated using fecal markers, and distal, but not proximal colonic pH<sup>[29]</sup>. Neither of these studies accounted for the known drift of the pH electrodes. The pH sensor drift is a function of time, therefore, studies in subjects with prolonged CTT tend to have a higher magnitude of pH change if drift corrections are not applied. In the present study we have examined the relationship between colonic pH and CTT measured using a radiotelemetry capsule (SmartPill) in healthy controls and constipated subjects.

# **METHODS**

#### **Subjects**

One hundred sixty eight subjects were enrolled in a study comparing CTT using the SmartPill wireless capsule manometry system versus radio opaque markers<sup>[30]</sup>. Eighty seven were healthy controls and 81 had chronic constipation based on Rome II criteria<sup>[31]</sup>. The healthy controls had no gastrointestinal symptoms or disorders when screened with the Mayo GI disease questionnaire<sup>[32]</sup>. Constipated patients were asked to discontinue all laxatives and drugs that affect motility at least 48 hours prior to the study. After an overnight fast subjects had a standardized meal and then ingested the SmartPill, which is a wireless motility capsule that measures pH, pressure and temperature; data are transmitted to a portable recorder and downloaded to a computer for analysis at the end of study<sup>[30]</sup>.

#### Measurement of CTT

CTT was measured as previously described by defining capsule entry into the cecum by a rapid pH drop of >1.0 unit, and capsule exit from the body by a sharp temperature drop and characteristic pressure signature<sup>[30]</sup>. Fifteen subjects, including 10 constipated and 5 healthy subjects, were excluded from analysis due to device malfunction or data loss. Five subjects, including 3 constipated and 2 controls, were excluded because of inability to define the ileocecal transition. Thus 148 subjects, 68 constipated and 80 healthy controls, were used for final analysis.

# Analysis of pH in different colonic segments

Mean pH was determined in time quartiles of colonic transit and in the fifteen minute intervals after entry into the cecum (proximal colon) and prior to body exit (distal colon) using the manufacturer's software. An experimentally determined drift correction of the SmartPill pH sensor of 0.01 pH units per hour was applied to the pH data. Constipated subjects were divided into normal and slow transit constipation based on the previously determined 95<sup>th</sup> percentile cutoff of 59 hours for the healthy controls<sup>[30]</sup>.

#### Statistical methods

To describe the observed variability in mean pH and test for differences between groups in this repeated measures design, a multivariate linear model was fit to the data. The endpoint was fit as a function of group, CTT quartile, and group by CTT quartile interaction. To account for the within-subject dependence structure, the model assumed that the distribution of the error terms for each subject to be multivariate normal with zero mean and an unstructured covariance structure. Once the model was fit, specific linear contrasts based on the estimated model parameters were constructed and used to test hypotheses of interest. Differences in univariate measures of pH were assessed using a standard ANOVA model. Spearman's correlation coefficient was used to assess the relationship between pH and CTT. All statistical analyses were carried out using SAS version 9.2 statistical software (Cary, NC).

### **RESULTS**

There was a weak, but significant correlation between CTT and pH in the distal colon. In the 15 minutes prior to body exit the correlation coefficient was R=0.20, p=0.0137, and in the fourth quartile of colonic transit time the correlation coefficient was R=0.22, p=0.006 (Figures 1, 2, table 1). No significant correlations between CTT and pH were found in the proximal colon (Table 1). The hydrogen ion concentration in the distal colon increased by 0.10 pH units for every 24 hour increase in colonic transit time.

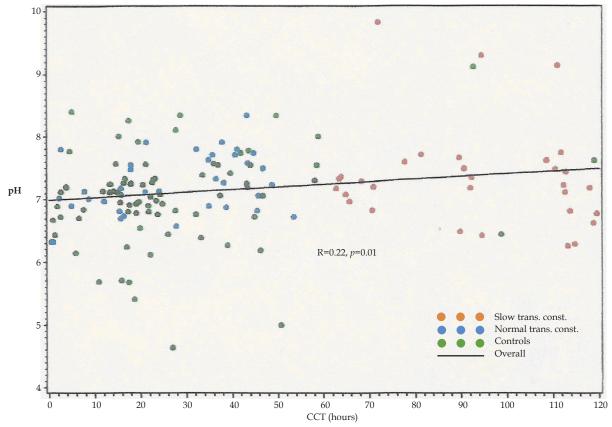


Figure 1 Correlation between CTT and mean colonic pH in the fourth quartile of colonic transit time.

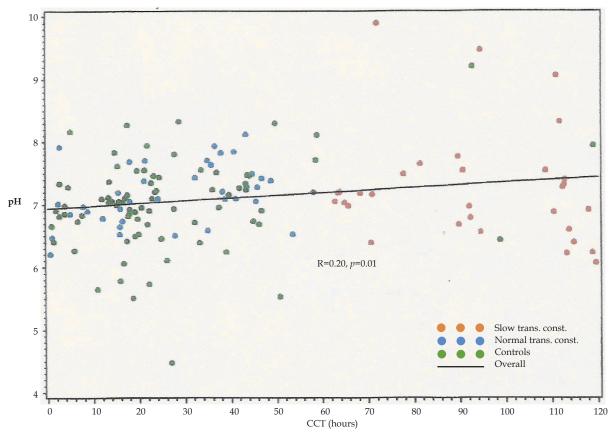


Figure 2 Correlation between CTT and mean colonic pH in the 15 mintues before capsule exit from the body.

<b>Table 1</b> Correlations between colonic transit time (CTT) and colonic pH in healthy controls and subjects with normal or slow transit constipation.					
	Controls	Normal transit constipation	Slow transit constipation	Correlation (R) between pH and CTT	P value
CTT (h)	27 (22)	27 (26)	90 (25)		
Quartile 1 pH	6.6 (0.5)	6.7 (0.5)	6.6 (0.5)	-0.04	0.54
Quartile 2 pH	6.8 (0.7)	7.0 (0.5)	7.0 (0.8)	0.03	0.67
Quartile 3 pH	7.0 (0.7)	7.3 (0.5)	7.3 (0.8)	0.14	0.08
Quarter 4 pH	7.1 (0.7)	7.2 (0.5)	7.4 (0.5)	0.22	0.01
15 min after cecal entry	6.5 (0.6)	6.7 (0.5)	6.5 (0.5)	0.08	0.36
15 min before body exit	7.0 (0.8)	7.2 (0.5)	7.3 (0.9)	0.20	0.01

There was an increase in the colonic pH during transit from the proximal (15 minutes after cecal entry) to the distal (15 minutes prior to body exit) colon in healthy controls and in subjects with either normal or slow transit constipation (Table 1). The mean rise in pH from the proximal to the distal colon tended to be greater in those with slow transit constipation (0.8 pH units) compared to normal controls (0.5 pH units) or to subjects with normal transit constipation (0.5 pH units) (p<0.09, slow transit constipation versus the other two groups).

# **DISCUSSION**

Few studies have addressed the effect of CTT on colonic pH. Lewis  $et~al^{[28]}$  used bran or drugs to alter whole gut transit, and examined the effects on colonic pH. Administration of wheat bran decreased whole gut transit from 69 hours to 39 hours (p=0.038), senna decreased transit from 69 hours to 41 hours (p=0.004), and loperamide increased whole gut transit from 50 hours to 74 hours (p=0.004). Distal colonic pH changed from baseline 7.08 to 6.88 in the wheat bran group (p=0.033), 7.14 to 6.66 in the senna group (p=0.039), and 7.11 to 7.15 in the loperamide group (p=0.80). Distal colonic pH was significantly associated with whole gut transit time (r=0.359, p=0.029) $^{[28]}$ . A study by Thomas  $et~al^{[29]}$  measured

colonic transit time and pH in patients with gallstones and stone-free controls. Patients with gallstones had longer CTT than stone-free controls (36.5 vs 23 hours) and higher distal colonic pH (7.67 vs 7.16). There was a linear relationship between CTT and distal colonic pH (r=0.62, p $\leq$ 0.001).

To our knowledge, the present study is the largest to examine the effect of CTT on colonic pH. We measured naturally occurring variations in pH and CTT in healthy controls, slow transit, and normal transit constipation subjects without manipulation by drugs or dietary changes. Alterations in fiber intake in the study by Lewis et al<sup>[28]</sup> could have affected colonic pH by affecting colonic bacterial SCFA production in addition to changing CTT. The CTT in the present study was measured simultaneously along with pH by a radiotelemetry device and not estimated from markers. In addition, data were corrected for the well-known time-dependent drift of pH electrodes, an important factor when assessing the impact of transit time on pH. Our results demonstrate a weak, but statistically significant correlation between CTT and pH in the distal, but not proximal colon. A rise in colonic pH from the cecum to the rectum was noted in all patient groups, and tended to be 0.3 pH units greater in those with slow transit constipation versus subjects with normal transit constipation or healthy controls.

Targeted delivery of orally administered drugs to the colon is often attempted by use of a pH sensitive coating, for example mesalamine preparations for treatment of ulcerative colitis. Targeted drug delivery systems utilizing pH coated preparations are also being studied for cancer chemotherapy<sup>[3]</sup>, treatment of colonic infectious agents<sup>[2]</sup>, and delivery of nutrients to colon<sup>[33]</sup>. Differences in distal colonic pH in patients caused by variations in CTT could significantly influence the dissolution of such preparations and the efficacy of drug delivery to the distal colon.

The colonic bacteria have important roles in human health. Some functions such as folate synthesis, generation of short chain fatty acids, and hydrogen sulfide production are well known. Potential influences of gut microbiota on metabolism and on the pathogenesis of obesity, diabetes, cancer, inflammatory bowel disease, and other disorders are being intensely examined<sup>[8]</sup>. A change in colonic pH of one unit or less has been shown to lead to alterations in species composition and metabolic output of the colonic bacteria<sup>[8]</sup>. For example, many Fermicutes are more tolerant of an acidic pH, giving them a competitive advantage<sup>[8]</sup>. Further studies are required to establish the impact of CTT and the resultant pH changes on colonic drug delivery and colonic physiology and pathophysiology.

# **ACKNOWLEDGMENTS**

This study was sponsored by the SmartPill Corporation and partly supported by the NYSTAR grant CO20118.

# CONFLICT OF INTERESTS

There are no conflicts of interest with regard to the present study.

### **REFERENCES**

- Yoshida T, Lai TC, Kwon GS, Sako K. pH- and ion-sensitive polymers for drug delivery. Expert Opinion on Drug Delivery 2013; 10(11): 1497-1513
- 2 Cassidy CM, Tunney MM, Caldwell DL, Andrews GP, Donnelly RF. Development of novel oral formulations prepared via hot melt extrusion for targeted delivery of photosensitizer to the colon. *Photochemistry and Photobiology* 2011; 87(4): 867-876
- 3 Sun G, Chang Y, Li S, Li Q, Xu R, Gu J, Wang E. pH-responsive controlled release of antitumour-active polyoxometalate from mesoporous silica materials. *Dalton Trans* 2009; 23: 4481-4487
- 4 McConnell EL, Liu F, Basit AW. Colonic treatments and targets: issues and opportunities. *J Drug Target* 2009; **17(5)**: 335-363
- 5 Chuong MC, Christensen JM, Ayres JW. Sustained delivery of intact drug to the colon: mesalamine formulation and temporal gastrointestinal transit analysis. *Pharm Dev Technol* 2009; **14(1)**: 116-125
- 6 Ibekwe VC, Fadda HM, McConnell EL, Khela MK, Evans DF, Basit AW. Interplay between intestinal pH, transit time and feed status on the in vivo performance of pH responsive ileo-colonic release systems. *Pharm Res* 2008; 25(8): 1828-1835
- Nugent SG, Kumar D, Rampton DS, Evans DF. Intestinal luminal pH in inflammatory bowel disease: possible determinants and implications for therapy with aminosalicylates and other drugs. *Gut* 2001; 48(4): 571-577
- 8 Flint HJ, Scott KP, Louis P, Duncan SH. The role of the gut microbiota in nutrition and health. *Nature Reviews Gastroenterology & Hepatology* 2012; 9(10): 577-589
- 9 Graff S, Chaumeil JC, Boy P, Lai-Kuen R, Charrueau C. Influence of pH conditions on the viability of Saccharomyces boulardii yeast. *J Gen Appl Microbiol* 2008; **54(4)**: 221-227
- 10 Flint HJ, Duncan SH, Scott KP, Louis P. Interactions and competition within the microbial community of the human colon: links

- between diet and health. Environ Microbiol 2007; 9(5): 1101-1111
  Duncan SH, Louis P, Flint HJ. Cultivable bacterial diversity from the human colon. *Lett Appl Microbiol* 2007; 44(4): 343-350
- 12 Walker AW, Duncan SH, McWilliam Leitch EC, Child MW, Flint HJ. pH and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. Appl Environ Microbiol 2005; 71(7): 3692-3700
- 13 Evans DF. Physicochemical environment of the colon. Eur J Cancer Prev 1998; 7 Suppl 2: S79-80
- 14 Swietach P, Patiar S, Supuran CT, Harris AL, Vaughan-Jones RD. The role of carbonic anhydrase 9 in regulating extracellular and intracellular ph in three-dimensional tumor cell growths. *J Biol Chem* 2009; 284(30): 20299-20310
- Bischof G, Cosentini E, Hamilton G, Riegler M, Zacherl J, Teleky B, Feil W, Schiessel R, Machen TE, Wenzl E. Effects of extracellular pH on intracellular pH-regulation and growth in a human colon carcinoma cell-line. *Biochim Biophys Acta* 1996; 1282(1): 131-139
- Said HM. Recent advances in transport of water-soluble vitamins in organs of the digestive system: a focus on the colon and the pancreas. Am J Physiol Gastrointest Liver Physiol 2013; 305(9): G601-610
- 17 Dudeja PK, Kode A, Alnounou M, Tyagi S, Torania S, Subramanian VS, Said HM. Mechanism of folate transport across the human colonic basolateral membrane. *Am J Physiol Gastrointest Liver Physiol* 2001; 281(1): G54-60
- 18 McDougall CJ, Wong R, Scudera P, Lesser M, DeCosse JJ. Colonic mucosal pH in humans. Dig Dis Sci 1993; 38(3): 542-545
- 19 Fallingborg J, Christensen LA, Ingeman-Nielsen M, Jacobsen BA, Abildgaard K, Rasmussen HH. pH-profile and regional transit times of the normal gut measured by a radiotelemetry device. *Ali*ment Pharmacol Ther 1989; 3(6): 605-613
- Evans DF, Pye G, Bramley R, Clark AG, Dyson TJ, Hardcastle JD. Measurement of gastrointestinal pH profiles in normal ambulant human subjects. *Gut* 1988; 29(8): 1035-1041
- 21 McNeil NI, Ling KL, Wager J. Mucosal surface pH of the large intestine of the rat and of normal and inflamed large intestine in man. Gut 1987; 28(6): 707-713
- 22 Zarate N, Mohammed SD, O'Shaughnessy E, Newell M, Yazaki E, Williams NS, Lunniss PJ, Semler JR, Scott SM. Accurate localization of a fall in pH within the ileocecal region: validation using a dual-scintigraphic technique. Am J Physiol Gastrointest Liver Physiol 2010; 299(6): G1276-1286
- 23 Macfarlane GT, Gibson GR, Cummings JH. Comparison of fermentation reactions in different regions of the human colon. J Appl Bacteriol 1992; 72(1): 57-64
- 24 Cummings JH. Fermentation in the human large intestine: evidence and implications for health. *Lancet* 1983; 1(8335): 1206-1209
- 25 Cummings JH, Macfarlane GT. The control and consequences of bacterial fermentation in the human colon. *J Appl Bacteriol* 1991; 70(6): 443-459
- 26 Miller TL, Wolin MJ. Fermentations by saccharolytic intestinal bacteria. Am J Clin Nutr 1979; 32(1): 164-172
- 27 Cook SI, Sellin JH. Review article: short chain fatty acids in health and disease. Aliment Pharmacol Ther 1998; 12(6): 499-507
- 28 Lewis SJ, Heaton KW. Increasing butyrate concentration in the distal colon by accelerating intestinal transit. *Gut* 1997; 41(2): 245-251
- 29 Thomas LA, Veysey MJ, Bathgate T, King A, French G, Smeeton NC, Murphy GM, Dowling RH. Mechanism for the transit-induced increase in colonic deoxycholic acid formation in cholesterol cholelithiasis. *Gastroenterology* 2000; 119(3): 806-815
- 30 Rao SS, Kuo B, McCallum RW, Chey WD, DiBaise JK, Hasler WL, Koch KL, Lackner JM, Miller C, Saad R, Semler JR, Sitrin MD, Wilding GE, Parkman HP. Investigation of colonic and whole-gut transit with wireless motility capsule and radiopaque

- markers in constipation. Clin Gastroenterol Hepatol 2009; 7(5): 537-544
- 31 Thompson WG, Longstreth GF, Drossman DA, Heaton KW, Irvine EJ, Muller-Lissner SA. Functional bowel disorders and functional abdominal pain. *Gut* 1999; **45 Suppl 2**: II43-47
- 32 Locke GR, Talley NJ, Weaver AL, Zinsmeister AR. A new questionnaire for gastroesophageal reflux disease. *Mayo Clin Proc* 1994; 69(6): 539-547
- 33 Aimone AM, Connolly B, Chaudhary S, Lugtu-Pe J, Martinuzzi F, Pencharz P, O'Connor DL. A combination of pH-sensitive caplet coatings may be an effective noninvasive strategy to deliver bioac-

tive substances, nutrients, or their precursors to the colon. *Applied Physiology, Nutrition, and Metabolism* 2009; **34(5)**: 893-900

Peer reviewers: Wei-Biao Cao, MD, Assistant Professor of Medicine, Department of Medicine & Pathology, Rhode Island Hospital and The Warren Alpert Medical School of Brown University, 55 Claverick St, Room 337, Providence, RI 02903, the United States; Sanda M. Cretoiu, MD, PhD, Division of Cellular and Molecular Medicine, Department of Morphological Sciences, Carol Davila University of Medicine and Pharmacy, 8 Eroii Sanitari Blvd., Bucharest 050474, Romania.